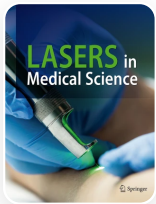


Home Lasers in Medical Science Article

Photobiomodulation enhances the Th1 immune response of human monocytes

Original Article Published: 06 November 2020

Volume 37, pages 135–148, (2022) Cite this article



Lasers in Medical Science

Aims and scope

Submit manuscript

Mayara Santos de Castro , Marta Miyazawa, Ester Siqueira Caixeta Nogueira, Jorge Kleber Chavasco, Gustavo Andrade Brancaglioni, Cláudio Daniel Cerdeira, Denismar Alves Nogueira, Marisa Ionta, João Adolfo Costa Hanemann, Maísa Ribeiro Pereira Lima Brigagão & Felipe Fornias Sperandio

Abstract

This study aims to evaluate the effects of photobiomodulation (PBM) on human monocytes, assessing the oxidative burst and ultimate fungicidal potential of

these cells, as well as the gene expression at the mRNA level of CD68, CD80, CD163, CD204, IL-6, TNF- α and IL-10 in derived macrophages. Primary cultures of human monocytes were irradiated with an InGaAlP (660 nm)/GaAlAs (780 nm) diode laser (parameters: 40 mW, 0.04 cm², 1 W/cm²; doses: 200, 400 and 600 J/cm²). Cells were submitted to the chemiluminescence assay, and a microbicidal activity assay against *Candida albicans* was performed. Reactive oxygen species (ROS) and nitric oxide (NO) production were measured, and cell viability was assessed by the exclusion method using 0.2% Trypan blue reagent. Irradiated monocytes were cultured for 72 h towards differentiation into macrophages. Total RNA was extracted, submitted to reverse transcription and real-time PCR. The results were analysed by ANOVA and the Tukey test ($\alpha = 0.05$). Irradiated monocytes revealed a significant increase in their intracellular and extracellular ROS ($P < 0.001$). The 660 nm wavelength and 400 J/cm² dose were the most relevant parameters ($P < 0.001$). The fungicidal capacity of the monocytes was shown to be greatly increased after PBM ($P < 0.001$). PBM increased the expression of TNF- α ($P = 0.0302$) and the production of NO ($P < 0.05$) and did not impair monocyte viability. PBM induces a pro-inflammatory Th1-driven response in monocytes and macrophages.

 This is a preview of subscription content, [log in via an institution](#)  to check access.

Access this article

[Log in via an institution](#)

Buy article PDF USD 39.95

Price excludes VAT (USA)

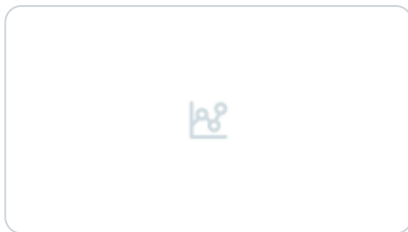
Tax calculation will be finalised during checkout.

Instant access to the full article PDF.

Rent this article via [DeepDyve](#) 

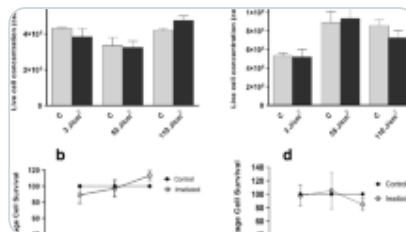
[Institutional subscriptions](#) →

Similar content being viewed by others



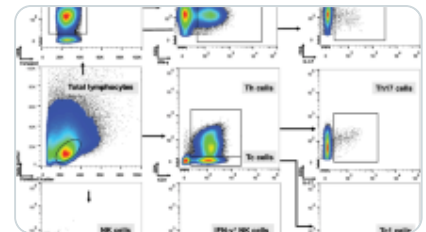
Violet/blue light activates Nrf2 signaling and modulates the inflammatory response...

Article | 01 June 2017



The viability of human cells irradiated with 470-nm light at various radiant energies *in vitro*

Article | 24 January 2021



Gallic acid diminishes pro-inflammatory interferon- γ - and interleukin-17-...

Article | Open access
09 February 2023

References

1. Cerdeira CD, Brigagão MRL, Carli ML, Ferreira CS, Moraes GOI, Hadad H, Hanemann JAC, Hamblin MR, Sperandio FF (2016) Low-level laser therapy stimulates the oxidative burst in human neutrophils and increases their fungicidal capacity. J Biophotonics 9:1180–1188.

<https://doi.org/10.1002/jbio.201600035>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

2. Eslamipour F, Motamedian SR, Bagheri F (2017) Ibuprofen and low-level laser therapy for pain control during fixed orthodontic therapy: a systematic review of randomized controlled trials and meta-analysis. J Contemp Dent Pract 18:527–533

[Article](#) [PubMed](#) [Google Scholar](#)

3. Heidari M, Paknejad M, Jamali R, Nokhbatolfoghahaei H, Fekrazad R, Moslemi N (2017) Effect of laser photobiomodulation on wound healing and postoperative pain following free gingival graft: a split-mouth triple-blind randomized controlled clinical trial. J Photochem Photobiol B 172:109–114.

<https://doi.org/10.1016/j.jphotobiol.2017.05.022>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

4. Karu T (1989) Photobiology of low-power laser effects. Health Phys 56:691–704

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

5. Hamblin MR (2017) Mechanisms and applications of the anti-inflammatory effects of photobiomodulation. AIMS Biophys 4:337–361.
<https://doi.org/10.3934/biophy.2017.3.337>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

6. Burger E, Mendes AC, Bani GM, Brigagão MR, Santos GB, Malaquias LC, Chavasco JK, Verinaud LM, de Camargo ZP, Hamblin MR, Sperandio FF (2015) Low-level laser therapy to the mouse femur enhances the fungicidal response of neutrophils against *Paracoccidioides brasiliensis*. PLoS Negl Trop Dis 9:e0003541. <https://doi.org/10.1371/journal.pntd.0003541>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

7. Hemvani N, Chitnis DS, Bhagwanani NS (2005) Helium–neon and nitrogen laser irradiation accelerates the phagocytic activity of human monocytes. Photomed Laser Surg 23:571–574. <https://doi.org/10.1089/pho.2005.23.571>

[Article](#) [PubMed](#) [Google Scholar](#)

8. Nathan CF, Root RK (1977) Hydrogen peroxide release from mouse peritoneal macrophages: dependence on sequential activation and triggering. J Exp Med 146:1648–1662

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

9. Dale DC, Boxer L, Liles WC (2008) The phagocytes: neutrophils and monocytes. Blood 112:935–945. <https://doi.org/10.1182/blood-2007>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

10. Rada B, Leto TL (2008) Oxidative innate immune defenses by Nox/Duox family NADPH oxidases. Contrib Microbiol 15:164–187. <https://doi.org/10.1159/000136357>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

11. Kalyanaraman B, Hardy M, Zielonka J (2016) A critical review of methodologies to detect reactive oxygen and nitrogen species stimulated by NADPH oxidase enzymes: implications in pesticide toxicity. Curr Pharmacol Rep 2:193–201. <https://doi.org/10.1007/s40495-016-0063-0>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

12. Liu H, Colavitti R, Rovira II, Finkel T (2005) Redox-dependent transcriptional regulation. Circ Res 97:967–974. <https://doi.org/10.1161/01.RES.0000188210.72062.10>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

13. Callaghan GA, Riordan C, Gilmore WS, McIntyre IA, Allen JM, Hannigan BM (1996) Reactive oxygen species inducible by low-intensity laser irradiation alter DNA synthesis in the haemopoietic cell line U937. *Lasers Surg Med* 19:201–206. [https://doi.org/10.1002/\(SICI\)1096-9101\(1996\)19:2<201::AID-LSM12>3.0.CO;2-9](https://doi.org/10.1002/(SICI)1096-9101(1996)19:2<201::AID-LSM12>3.0.CO;2-9)

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

14. Lindgård A, Hultén LM, Svensson L, Soussi B (2007) Irradiation at 634 nm releases nitric oxide from human monocytes. *Lasers Med Sci* 22:30–36. <https://doi.org/10.1007/s10103-006-0419-5>

[Article](#) [PubMed](#) [Google Scholar](#)

15. Silva IH, de Andrade SC, de Faria AB, Fonsêca DD, Gueiros LA, Carvalho AA, da Silva WT, de Castro RM, Leão JC (2016) Increase in the nitric oxide release without changes in cell viability of macrophages after laser therapy with 660 and 808 nm lasers. *Lasers Med Sci* 31:1855–1862. <https://doi.org/10.1007/s10103-016-2061-1>

[Article](#) [PubMed](#) [Google Scholar](#)

16. Chen CH, Wang CZ, Wang YH, Liao WT, Chen YJ, Kuo CH, Kuo HF, Hung CH (2014) Effects of low-level laser therapy on M1-related cytokine expression in monocytes via histone modification. *Mediat Inflamm* 2014:625048. <https://doi.org/10.1155/2014/625048>

[Article](#) [CAS](#) [Google Scholar](#)

17. de Brito Sousa K, Rodrigues MFSD, de Souza Santos D, Mesquita-Ferrari RA, Nunes FD, da Silva DFTDifferential expression of inflammatory and anti-inflammatory mediators by M1 and M2 macrophages after photobiomodulation with red or infrared lasers, Bussadori SK, Fernandes KPS (2019) . Lasers Med Sci <https://doi.org/10.1007/s10103-019-02817-1>

18. Yin M, Shen J, Yu S, Fei J, Zhu X, Zhao J, Zhai L, Sadhukhan A, Zhou J (2019) Tumor-associated macrophages (TAMs): a critical activator in ovarian cancer metastasis. Onco Targets Ther 12:8687–8699. <https://doi.org/10.2147/OTT.S216355>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

19. Tarique AA, Logan J, Thomas E, Holt PG, Sly PD, Fantino E (2015) Phenotypic, functional, and plasticity features of classical and alternatively activated human macrophages. Am J Respir Cell Mol Biol 53:676–688. <https://doi.org/10.1165/rcmb.2015-0012OC>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

20. Yaniv E, Hadar T, Shvero J, Tamir R, Nageris B (2009) KTP/532 YAG laser treatment for allergic rhinitis. Am J Rhinol Allergy 23:527–530. <https://doi.org/10.2500/ajra.2009.23.3346>

[Article](#) [PubMed](#) [Google Scholar](#)

21. Fairweather D, Cihakova D (2009) Alternatively activated macrophages in infection and autoimmunity. *J Autoimmun* 33:222–230.

<https://doi.org/10.1016/j.jaut.2009.09.012>

[Article](#) [CAS](#) [Google Scholar](#)

22. Hultén LM, Holmström M, Soussi B (1999) Harmful singlet oxygen can be helpful. *Free Radic Biol Med* 27:1203–1207

[Article](#) [PubMed](#) [Google Scholar](#)

23. Babior BM (1984) The respiratory burst of phagocytes. *J Clin Invest* 73:599–601. <https://doi.org/10.1172/JCI111249>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

24. Dahlgren C, Karlsson A, Bylund J (2007) Measurement of respiratory burst products generated by professional phagocytes. *Methods Mol Biol* 412:349–363. https://doi.org/10.1007/978-1-59745-467-4_23

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

25. Silva DF, Mesquita-Ferrari RA, Fernandes KP, Raele MP, Wetter NU, Deana AM (2012) Effective transmission of light for media culture, plates and tubes. *Photochem Photobiol* 88:1211–1216. <https://doi.org/10.1111/j.1751-1097.2012.01166.x>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

26. Green JN, Winterbourn CC, Hampton MB (2007) Analysis of neutrophil bactericidal activity. *Methods Mol Biol* 412:319–332.

https://doi.org/10.1007/978-1-62703-845-4_19

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

27. Raggi F, Pelassa S, Pierobon D, Penco F, Gattorno M, Novelli F, Eva A, Varesio L, Giovarelli M, Bosco MC (2017) Regulation of human macrophage M1-M2 polarization balance by hypoxia and the triggering receptor expressed on myeloid cells-1. *Front Immunol* 8:1097.

<https://doi.org/10.3389/fimmu.2017.01097>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

28. Jaguin M, Houlbert N, Fardel O, Lecureur V (2013) Polarization profiles of human M-CSF-generated macrophages and comparison of M1-markers in classically activated macrophages from GM-CSF and M-CSF origin. *Cell Immunol* 281:51–61. <https://doi.org/10.1016/j.cellimm.2013.01.010>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

29. Meirelles MN, Araujo TCJ, Souza W (1980) Interaction of epimastigote and trypomastigote forms of *Trypanosoma cruzi* with chicken macrophages in vitro. *Parasitology* 81:373–381

[Article](#) [PubMed](#) [Google Scholar](#)

30. Ding J, Jin W, Chen C, Shao Z, Wu J (2012) Tumor associated macrophage × cancer cell hybrids may acquire cancer stem cell properties in breast cancer. PLoS One 7:e41942. <https://doi.org/10.1371/journal.pone.0041942>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

31. Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paepe A, Speleman F (2002) Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. Genome Biol 3:RESEARCH0034

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

32. Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta C(T)}$ method. Method Methods 25:402–408. <https://doi.org/10.1006/meth.2001.1262>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

33. R Core Team (2016) R Foundation for Statistical Computing

34. Ferreira EB, Cavalcanti PP, Nogueira DA (2013) R package version:1.1.2

35. Pal G, Dutta A, Mitra K, Grace MS, Romanczyk TB, Wu X, Chakrabarti K, Anders J, Gorman E, Waynant RW, Tata DB (2007) Effect of low intensity laser interaction with human skin fibroblast cells using fiber-optic nano-probes. J Photochem Photobiol B 86:252–261.

<https://doi.org/10.1016/j.jphotobiol.2006.12.001>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

36. Huang YY, Chen AC, Carroll JD, Hamblin MR (2009) Biphasic dose response in low level light therapy. Dose-Response 7:358–383.

<https://doi.org/10.2203/dose-response.09-027.Hamblin>

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

37. Huang YY, Sharma SK, Carroll J, Hamblin MR (2011) Biphasic dose response in low level light therapy - an update. Dose-Response 9:602–618.

<https://doi.org/10.2203/dose-response.11-009.Hamblin>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

38. Ellis JA, Mayer SJ, Jones OT (1988) The effect of the NADPH oxidase inhibitor diphenyleneiodonium on aerobic and anaerobic microbicidal activities of human neutrophils. Biochem J 251:887–891.

<https://doi.org/10.1042/bj2510887>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

39. Tucureanu MM, Rebleanu D, Constantinescu CA, Deleanu M, Voicu G, Butoi E, Calin M, Manduteanu I (2017) Lipopolysaccharide-induced inflammation in monocytes/macrophages is blocked by liposomal delivery of Gi-protein inhibitor. *Int J Nanomedicine* 13:63–76.

<https://doi.org/10.2147/IJN.S150918>

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

40. Akira S, Takeda K (2004) Toll-like receptor signalling. *Nat Rev Immunol* 4:499–511. <https://doi.org/10.1038/nri1391>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

41. Raetz CR, Whitfield C (2002) Lipopolysaccharide endotoxins. *Annu Rev Biochem* 71:635–700.

<https://doi.org/10.1146/annurev.biochem.71.110601.135414>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

42. Rossol M, Heine H, Meusch U, Quandt D, Klein C, Sweet MJ, Hauschildt S (2011) LPS-induced cytokine production in human monocytes and macrophages. *Crit Rev Immunol* 31:379–446

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

43. Netea MG, Suttmuller R, Hermann C, Van der Graaf CA, Van der Meer JW, van Krieken JH, Hartung T, Adema G, Kullberg BJ (2004) Toll-like receptor 2 suppresses immunity against *Candida albicans* through induction of IL-10 and regulatory T cells. *J Immunol* 172:3712–3718

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

44. Netea MG, Van der Graaf C, Van der Meer JW, Kullberg BJ (2004) Recognition of fungal pathogens by toll-like receptors. *Eur J Clin Microbiol Infect Dis* 23:672–676. <https://doi.org/10.1007/s10096-004-1192-7>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

45. Farah CS, Saunus JM, Hu Y, Kazoullis A, Ashman RB (2009) Gene targeting demonstrates that inducible nitric oxide synthase is not essential for resistance to oral candidiasis in mice, or for killing of *Candida albicans* by macrophages in vitro. *Oral Microbiol Immunol* 24:83–88. <https://doi.org/10.1111/j.1399-302X.2008.00462.x>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

46. Avci P, Gupta A, Sadasivam M, Vecchio D, Pam Z, Pam N, Hamblin MR (2013) Low-level laser (light) therapy (LLLT) in skin: stimulating, healing, restoring. *Semin Cutan Med Surg* 32:41–52

[PubMed](#) [PubMed Central](#) [Google Scholar](#)

47. Sperandio FF, Simoes A, Aranha AC, Correa L, Orsini Machado de Sousa SC (2010) Photodynamic therapy mediated by methylene blue dye in wound healing. *Photomed Laser Surg* 28:581–587.
<https://doi.org/10.1089/pho.2009.2601>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

48. Sperandio FF, Simoes A, Correa L, Aranha AC, Giudice FS, Hamblin MR, Sousa SC (2015) Low-level laser irradiation promotes the proliferation and maturation of keratinocytes during epithelial wound repair. *J Biophotonics* 8:795–803. <https://doi.org/10.1002/jbio.201400064>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

49. Viegas VN, Abreu ME, Viezzer C, Machado DC, Filho MS, Silva DN, Pagnoncelli RM (2007) Effect of low-level laser therapy on inflammatory reactions during wound healing: comparison with meloxicam. *Photomed Laser Surg* 25:467–473. <https://doi.org/10.1089/pho.2007.1098>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

50. Woodruff LD, Bounkeo JM, Brannon WM, Dawes KS, Barham CD, Waddell DL, Enwemeka CS (2004) The efficacy of laser therapy in wound repair: a meta-analysis of the literature. *Photomed Laser Surg* 22:241–247.
<https://doi.org/10.1089/1549541041438623>

[Article](#) [PubMed](#) [Google Scholar](#)

51. Moriyama Y, Nguyen J, Akens M, Moriyama EH, Lilge L (2009) In vivo effects of low level laser therapy on inducible nitric oxide synthase. *Lasers Surg Med* 41:227–231. <https://doi.org/10.1002/lsm.20745>

[Article](#) [PubMed](#) [Google Scholar](#)

52. Labonte AC, Tosello-Tramont AC, Hahn YS (2014) The role of macrophage polarization in infectious and inflammatory diseases. *Mol Cell* 37:275–285. <https://doi.org/10.14348/molcells.2014.2374>

[Article](#) [CAS](#) [Google Scholar](#)

53. Sica A, Mantovani A (2012) Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 122:787–795. <https://doi.org/10.1172/JCI59643>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

54. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M (2004) The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* 25:677–686. <https://doi.org/10.1016/j.it.2004.09.015>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

55. Martinez FO, Helming L, Gordon S (2009) Alternative activation of macrophages: an immunologic functional perspective. *Annu Rev Immunol* 27:451–483. <https://doi.org/10.1146/annurev.immunol.021908.132532>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

56. Colvin EK (2014) Tumor-associated macrophages contribute to tumor progression in ovarian cancer. *Front Oncol* 4:137. <https://doi.org/10.3389/fonc.2014.00137>

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

57. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A (2002) Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* 23:549–555. [https://doi.org/10.1016/s1471-4906\(02\)02302-5](https://doi.org/10.1016/s1471-4906(02)02302-5)

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

58. Homan JW, Steele AD, Martinand-Mari C, Rogers TJ, Henderson EE, Charubala R, Pfeleiderer W, Reichenbach NL, Suhadolnik RJ (2002) Inhibition of morphine-potentiated HIV-1 replication in peripheral blood mononuclear cells with the nuclease-resistant 2-5A agonist analog, 2-5A(N6B). *J Acquir Immune Defic Syndr* 30:9–20. <https://doi.org/10.1097/00042560-200205010-00002>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

59. Lagging M, Romero AI, Westin J, Norkrans G, Dhillon AP, Pawlotsky JM, Zeuzem S, von Wagner M, Negro F, Schalm SW, Haagmans BL, Ferrari C, Missale G, Neumann AU, Verheij-Hart E, Hellstrand K (2006) IP-10 predicts viral response and therapeutic outcome in difficult-to-treat patients with HCV genotype 1 infection. *Hepatology* 44:1617–1625.
<https://doi.org/10.1002/hep.21407>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

Acknowledgements

The authors wish to thank Dr. Janete M. Cerutti for some reagents provided for the molecular study.

Funding

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) (Master's Scholarship).

Author information

Authors and Affiliations

Department of Pathology and Parasitology, Institute of Biomedical Sciences, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil

Mayara Santos de Castro & Felipe Fornias Sperandio

Department of Clinic and Surgery, School of Dentistry, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Marta Miyazawa & João Adolfo Costa Hanemann

Department of Cell and Developmental Biology, Institute of Biomedical Sciences, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Ester Siqueira Caixeta Nogueira & Marisa Ionta

Department of Microbiology and Immunology, Institute of Biomedical Sciences, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Jorge Kleber Chavasco

Central Analysis Laboratory (LACEN), Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Gustavo Andrade Brancaglioni

Department of Biochemistry, Institute of Biomedical Sciences, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Cláudio Daniel Cerdeira & Máisa Ribeiro Pereira Lima Brigagão

Institute of Exact Sciences, Federal University of Alfenas, 700 Gabriel Monteiro da Silva Street, Alfenas, MG, 37130-000, Brazil
Denismar Alves Nogueira

Oral Medicine Oral Pathology Resident – Faculty of Dentistry, University of British Columbia, 2199 Wesbrook Mall, Vancouver, BC, V6T 1Z3, Canada
Felipe Fornias Sperandio

Corresponding author

Correspondence to [Mayara Santos de Castro](#).

Ethics declarations

Conflict of interest

The authors declare that they have no conflict of interest.

Ethical approval

The present study was approved by the Human Research Ethics Committee of the University Federal of Alfenas, Brazil (Protocol # 1980248).

Informed consent

Informed consent was obtained from all individual participants included in the study.

Additional information

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary information

[Fig. S1](#)

The process of differentiating monocytes into macrophages was followed for 72 h, observing the change in the morphological aspect of the cells. Monocytes (arrow) have a rounded morphology, whilst macrophages (star) cease to be rounded cells and become elongated, spindle shaped and often stellar. Image obtained by inverted microscopy in phase contrast ($\times 400$). (PNG 772 kb)

[High resolution \(TIF 1332 kb\)](#)

[Table S1](#)

(DOCX 36 kb)

Rights and permissions

[Reprints and permissions](#)

About this article

Cite this article

de Castro, M.S., Miyazawa, M., Nogueira, E.S.C. *et al.* Photobiomodulation enhances the Th1 immune response of human monocytes. *Lasers Med Sci* **37**, 135–148 (2022).

<https://doi.org/10.1007/s10103-020-03179-9>

Received

10 August 2020

Accepted

27 October 2020

Published

06 November 2020

Issue Date

February 2022

DOI

<https://doi.org/10.1007/s10103-020-03179-9>

Keywords

[Photobiomodulation](#)

[Monocytes](#)

[Macrophages](#)

[Candida albicans](#)

[Reactive oxygen species](#)

[Th1 immune response](#)